

Toxo IgM

IgM antibodies to *Toxoplasma gondii*

cobas®

REF	Σ	SYSTEM
04618858 190	100	Elecsys 2010 MODULAR ANALYTICS E170 cobas e 411 cobas e 601 cobas e 602

English

Intended use

Immunoassay for the in vitro qualitative determination of IgM antibodies to *Toxoplasma gondii* in human serum and plasma.

The electrochemiluminescence immunoassay "ECLIA" is intended for use on Elecsys and **cobas e** immunoassay analyzers.

Summary

Toxoplasmosis is a common infection caused by the protozoan parasite *Toxoplasma gondii*.

The infection is mainly acquired by ingestion of food or water that is contaminated by mature oocysts shed by cats or by undercooked meat containing tissue cysts.¹

Primary, acute infection which is mostly mild or even asymptomatic in healthy individuals, is followed by a latent infection which usually persists for life. However, reactivation of a latent *Toxoplasma* infection as a result of immunosuppression (e.g. in organ transplant recipients, AIDS patients) is frequently associated with meningoencephalitis.^{2,3}

Primary maternal *Toxoplasma* infection occurring during pregnancy may lead to severe damage of the fetus as the parasite can be transmitted across the placenta. The majority of infants with congenital infection do not present clinical symptoms at birth but may develop severe sequelae later in life like mental and psychomotor retardation, chorioretinitis and hearing loss.⁴ The fetal infection rate increases with gestational age. However, the risk of severe clinical manifestations is higher in case of early maternal infection.^{4,5,6}

Early drug therapy in acute infection during pregnancy can prevent congenital damage or ameliorate the severity of clinical manifestations.^{4,5,6}

The diagnosis of *Toxoplasma* infection is most commonly made by the detection of anti-*Toxoplasma*-specific IgG and IgM antibodies.

The detection of Toxo IgM antibodies is presumptive of an acute, recent or reactivated *Toxoplasma* infection.

The determination of IgG antibodies to *Toxoplasma gondii* is used to assess the serological status to *T. gondii* and is indicative of an acute or latent infection.

The diagnosis of the acute acquired infection during pregnancy is established by a seroconversion or a significant rise in antibody titers (IgG and/or IgM) in serial samples.^{4,6}

Test principle

μ -Capture test principle. Total duration of assay: 18 minutes.

- 1st incubation: 10 μ L of sample are automatically prediluted 1:20 with Diluent Universal. *T. gondii*-specific recombinant antigen labeled with a ruthenium complex^{a)} is added. Anti-Toxo IgM antibodies present in the sample react with the ruthenium-labeled *T. gondii*-specific recombinant antigen.
- 2nd incubation: Biotinylated monoclonal h-IgM-specific antibodies and streptavidin-coated microparticles are added. The complex becomes bound to the solid phase via interaction of biotin and streptavidin.
- The reaction mixture is aspirated into the measuring cell where the microparticles are magnetically captured onto the surface of the electrode. Unbound substances are then removed with ProCell/ProCell M. Application of a voltage to the electrode then induces chemiluminescent emission which is measured by a photomultiplier.
- Results are determined automatically by the software by comparing the electrochemiluminescence signal obtained from the reaction product of the sample with the signal of the cutoff value previously obtained by calibration.

a) Tris(2,2'-bipyridyl)ruthenium(II)-complex (Ru(bpy)₃²⁺)

Reagents - working solutions

The reagent rackpack (M, R1, R2) is labeled as TOXIGM.

- M Streptavidin-coated microparticles (transparent cap), 1 bottle, 6.5 mL: Streptavidin-coated microparticles 0.72 mg/mL; preservative.
- R1 *Toxoplasma*-Ag-Ru(bpy)₃²⁺ (gray cap), 1 bottle, 9 mL: *Toxoplasma*-antigen labeled with ruthenium complex > 1 mg/L; MES^{b)} buffer 50 mmol/L, pH 6.0; preservative.
- R2 Anti-h-IgM-Ab-biotin (black cap), 1 bottle, 9 mL: Biotinylated monoclonal anti-h-IgM antibody (mouse) > 500 μ g/L; HEPES^{c)} buffer 50 mmol/L, pH 7.2; preservative.

b) MES = 2-morpholino-ethane sulfonic acid

c) HEPES = [4-(2-hydroxyethyl)-piperazine]-ethane sulfonic acid

- TOXIGM Cal1 Negative calibrator 1 (white cap), 2 bottles of 0.67 mL each:
Human serum, negative for anti-Toxo IgM; preservative.
- TOXIGM Cal2 Positive calibrator 2 (black cap), 2 bottles of 0.67 mL each:
Anti-Toxo IgM (human) approximately 130 U/mL (Roche units) in human serum; preservative.

Precautions and warnings

For in vitro diagnostic use.

Exercise the normal precautions required for handling all laboratory reagents.

Disposal of all waste material should be in accordance with local guidelines. Safety data sheet available for professional user on request.

All human material should be considered potentially infectious. All products derived from human blood (TOXIGM Cal1, TOXIGM Cal2) are prepared exclusively from the blood of donors tested individually and shown to be free from HBsAg and antibodies to HCV and HIV.

The serum containing anti-Toxo IgM (TOXIGM Cal2) was sterile filtrated.

The testing methods applied were FDA-approved or cleared in compliance with the European Directive 98/79/EC, Annex II, List A.

However, as no testing method can rule out the potential risk of infection with absolute certainty, the material should be handled with the same level of care as a patient specimen. In the event of exposure, the directives of the responsible health authorities should be followed.^{7,8}

Avoid foam formation in all reagents and sample types (specimens, calibrators and controls).

Reagent handling

The reagents in the kit are ready for use and are supplied in bottles compatible with the system.

Elecsys 2010 and **cobas e 411** analyzers: The calibrators should only be left on the analyzers during calibration at 20-25 °C. After use, close the bottles as soon as possible and store upright at 2-8 °C.

Due to possible evaporation effects, not more than 5 calibration procedures per bottle set should be performed.

MODULAR ANALYTICS E170, **cobas e 601** and **cobas e 602** analyzers: Unless the entire volume is necessary for calibration on the analyzer, transfer aliquots of the ready-for-use calibrators into empty snap-cap bottles (CalSet Vials). Attach the supplied labels to these additional bottles. Store the aliquots at 2-8 °C for later use.

Perform **only one** calibration procedure per aliquot.

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All information required for correct operation is read in from the respective reagent barcodes.

Please note: Both the vial labels, and the additional labels (if available) contain 2 different barcodes. The barcode between the yellow markers is for **cobas** 8000 systems only. If using a **cobas** 8000 system, please turn the vial cap 180° into the correct position so the barcode can be read by the system. Place the vial on the instrument as usual.

Storage and stability

Store at 2-8 °C.

Do not freeze.

Store the Elecsys reagent kit **upright** in order to ensure complete availability of the microparticles during automatic mixing prior to use.

Stability of the reagent rackpack	
unopened at 2-8 °C	up to the stated expiration date
after opening at 2-8 °C	12 weeks
on the analyzers	2 weeks or 12 weeks if stored alternately in the refrigerator and on the analyzers (up to 84 hours)

Stability of the calibrators	
unopened at 2-8 °C	up to the stated expiration date
after opening at 2-8 °C	8 weeks
on Elecsys 2010 and cobas e 411 at 20-25 °C	up to 5 hours
on MODULAR ANALYTICS E170, cobas e 601 and cobas e 602 at 20-25 °C	use only once

Store calibrators **upright** in order to prevent the calibrator solution from adhering to the snap-cap.

Specimen collection and preparation

Only the specimens listed below were tested in a sufficient number and found acceptable.

Serum collected using standard sampling tubes or tubes containing separating gel.

Li-heparin, K₃-EDTA and Na-citrate plasma.

Criterion: Mean recovery of positive samples within 80-120 % of serum value.

Stable for 3 weeks at 2-8 °C, 3 days at 25 °C, 3 months at -20 °C. The samples may be frozen 6 times.

The sample types listed were tested with a selection of sample collection tubes that were commercially available at the time of testing, i.e. not all available tubes of all manufacturers were tested. Sample collection systems from various manufacturers may contain differing materials which could affect the test results in some cases. When processing samples in primary tubes (sample collection systems), follow the instructions of the tube manufacturer.

Specimens should not be subsequently altered with additives (biocides, anti-oxidants or substances that could possibly change the pH of the sample) in order to avoid erroneous findings.

Pooled samples and other artificial material may have different effects on different assays and thus may lead to discrepant findings.

Centrifuge samples containing precipitates and frozen samples before performing the assay. Lyophilized samples, heat-inactivated samples and samples and controls stabilized with azide (up to 1 %) can be used.

Ensure the samples, calibrators and controls are at 20-25 °C prior to measurement.

Due to possible evaporation effects, samples, calibrators and controls on the analyzers should be analyzed/measured within 2 hours.

Materials provided

See "Reagents – working solutions" section for reagents.

- 2 x 2 bottle labels

Materials required (but not provided)

- [REF]** 04618866190, PreciControl Toxo IgM, 8 x 0.67 mL each of PreciControl Toxo IgM 1 and 2
 - [REF]** 11732277122, Diluent Universal, 2 x 16 mL sample diluent or **[REF]** 03183971122, Diluent Universal, 2 x 36 mL sample diluent
 - [REF]** 11776576322, CalSet Vials, 2 x 56 empty snap-cap bottles
 - General laboratory equipment
 - Elecsys 2010, MODULAR ANALYTICS E170 or **cobas e** analyzer
- Accessories for Elecsys 2010 and **cobas e** 411 analyzers:
- [REF]** 11662988122, ProCell, 6 x 380 mL system buffer
 - [REF]** 11662970122, CleanCell, 6 x 380 mL measuring cell cleaning solution
 - [REF]** 11930346122, Elecsys SysWash, 1 x 500 mL washwater additive
 - [REF]** 11933159001, Adapter for SysClean
 - [REF]** 11706802001, Elecsys 2010 AssayCup, 60 x 60 reaction vessels
 - [REF]** 11706799001, Elecsys 2010 AssayTip, 30 x 120 pipette tips
- Accessories for MODULAR ANALYTICS E170, **cobas e** 601 and **cobas e** 602 analyzers:

- [REF]** 04880340190, ProCell M, 2 x 2 L system buffer
 - [REF]** 04880293190, CleanCell M, 2 x 2 L measuring cell cleaning solution
 - [REF]** 03023141001, PC/CC-Cups, 12 cups to prewarm ProCell M and CleanCell M before use
 - [REF]** 03005712190, ProbeWash M, 12 x 70 mL cleaning solution for run finalization and rinsing during reagent change
 - [REF]** 12102137001, AssayTip/AssayCup Combimagazine M, 48 magazines x 84 reaction vessels or pipette tips, waste bags
 - [REF]** 03023150001, WasteLiner, waste bags
 - [REF]** 03027651001, SysClean Adapter M
- Accessories for all analyzers:
- [REF]** 11298500316, ISE Cleaning Solution/Elecsys SysClean, 5 x 100 mL system cleaning solution

Assay

For optimum performance of the assay follow the directions given in this document for the analyzer concerned. Refer to the appropriate operator's manual for analyzer-specific assay instructions.

Resuspension of the microparticles takes place automatically prior to use. Read in the test-specific parameters via the reagent barcode. If in exceptional cases the barcode cannot be read, enter the 15-digit sequence of numbers.

Bring the cooled reagents to approximately 20 °C and place on the reagent disk (20 °C) of the analyzer. Avoid foam formation. The system automatically regulates the temperature of the reagents and the opening/closing of the bottles.

Place the calibrators in the sample zone.

All the information necessary for calibrating the assay is automatically read into the analyzer.

After calibration has been performed, store the calibrators at 2-8 °C or discard (MODULAR ANALYTICS E170, **cobas e** 601 and **cobas e** 602 analyzers).

Calibration

Traceability: This method has been standardized against a Roche standard. The units have been selected arbitrarily.

Calibration frequency: Calibration must be performed once per reagent lot using TOXIGM Cal1, TOXIGM Cal2 and fresh reagent (i.e. not more than 24 hours since the reagent kit was registered on the analyzer). Renewed calibration is recommended as follows:

- after 1 month (28 days) when using the same reagent lot
- after 7 days (when using the same reagent kit on the analyzer)
- as required: e.g. quality control findings outside the defined limits
- more frequently when this is required by pertinent regulations

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Range for the electrochemiluminescence signals (counts) for the calibrators:

Negative calibrator (TOXIGM Cal1): 400-2500 (Elecsys 2010, MODULAR ANALYTICS E170 and **cobas e** analyzers).

Positive calibrator (TOXIGM Cal2): 4500-35000 (Elecsys 2010, MODULAR ANALYTICS E170 and **cobas e** analyzers).

Quality control

For quality control, use PreciControl Toxo IgM.

Controls for the various concentration ranges should be run individually at least once every 24 hours when the test is in use, once per reagent kit, and following each calibration.

The control intervals and limits should be adapted to each laboratory's individual requirements. Values obtained should fall within the defined limits. Each laboratory should establish corrective measures to be taken if values fall outside the defined limits.

If necessary, repeat the measurement of the samples concerned.

Follow the applicable government regulations and local guidelines for quality control.

Note:

For technical reasons re-assigned target values valid only for a specific reagent and control lot combination, must be entered manually on all analyzers (except for the **cobas e** 602 analyzer). Therefore always refer to the value sheet included in the rackpack or PreciControl kit to make sure that the correct target values are used.

When a new reagent or control lot is used, the analyzer will use the original values encoded in the control barcodes.

Calculation

The analyzer automatically calculates the cutoff based on the measurement of TOXIGM Cal1 and TOXIGM Cal2. The result of a sample is given either as reactive or non-reactive as well as in the form of a cutoff index (signal sample/cutoff).

Interpretation of the results

Results obtained with the Elecsys Toxo IgM assay can be interpreted as follows:

Non-reactive: < 0.8 COI

Indeterminate: ≥ 0.8 - < 1.0 COI

Reactive: ≥ 1.0 COI

Samples with a cutoff index < 0.8 are non-reactive in the Elecsys Toxo IgM assay.

Samples with a cutoff index between ≥ 0.8 and < 1.0 are considered indeterminate. The sample should be retested. In case the result is still indeterminate, a second sample should be tested e.g. within 2-3 weeks. Samples with a cutoff index ≥ 1.0 are reactive in the Elecsys Toxo IgM assay.

The magnitude of the measured result above the cutoff is not indicative of the total amount of antibody present in the sample.

The anti-Toxoplasma IgM results in a given specimen, as determined by assays from different manufacturers, can vary due to differences in assay and reagent methods.

Limitations - interference

A negative Toxo IgM test result, also in combination with a positive Toxo IgG result, does not completely rule out the possibility of an acute infection with *Toxoplasma gondii*:

- Individuals at the early stage of acute infection may not exhibit detectable amounts of Toxo IgM antibodies. In some of these individuals an indeterminate or low positive result with the Elecsys Toxo IgG assay may be found and indicate an early acute infection. A second sample should be tested e.g. within 2 weeks. The detection of Toxo IgM and/or a significant increase of the Elecsys Toxo IgG antibody titer in the second sample supports the diagnosis of acute Toxoplasma infection.
- In some individuals Toxoplasma IgM-specific antibodies may revert to non-reactive levels within few weeks after infection with *T. gondii*.

The detection of IgM antibodies against *T. gondii* in a single sample is not sufficient to prove an acute Toxoplasma infection since elevated IgM antibody levels may persist even for years after initial infection.^{9,10} Further tests or a combination of test methods should be done for clarification.^{1,4,5,10} A significant increase of the Toxo IgG antibody titer from a first to a second

sample taken e.g. within 2 weeks may support the diagnosis of acute Toxoplasma infection.

If a treatment is prescribed early enough, antibody production may not increase. IgG and IgM levels may remain low and can coexist for years.

Elecsys Toxo IgM results should be used in conjunction with Toxoplasma-specific IgG results, the patient's medical history, clinical symptoms and other laboratory tests.

The results in HIV patients, in patients undergoing immunosuppressive therapy, or in patients with other disorders leading to immune suppression, should be interpreted with caution.

Specimens from neonates, cord blood, pretransplant patients or body fluids other than serum and plasma, such as urine, saliva or amniotic fluid have not been tested.

The assay is unaffected by icterus (bilirubin < 684 µmol/L or < 40 mg/dL), hemolysis (Hb < 1.24 mmol/L or < 2 g/dL), lipemia (Intralipid < 2000 mg/dL) and biotin (< 246 nmol/L or < 60 ng/mL).

Criterion: Mean recovery of positive samples within ± 20 % of serum value.

Samples should not be taken from patients receiving therapy with high biotin doses (i.e. > 5 mg/day) until at least 8 hours following the last biotin administration.

No interference was observed from rheumatoid factors up to a concentration of 3720 IU/mL.

The high-dose hook effect does not lead to false-negative results in the Elecsys Toxo IgM assay.

In vitro tests were performed on 18 commonly used pharmaceuticals and in addition on spiramycin, sulfadiazine, folic acid and pyrimethamine. No interference with the assay was found.

As with many µ-capture assays an interference with unspecific IgM is observed. Increasing amounts of unspecific IgM may lead to a decrease in the recovery of positive samples with the Elecsys Toxo IgM assay.

In rare cases, interference due to extremely high titers of antibodies to immunological components, streptavidin or ruthenium can occur. These effects are minimized by suitable test design.

For diagnostic purposes, the results should always be assessed in conjunction with the patient's medical history, clinical examination and other findings.

Specific performance data

Representative performance data on the analyzers are given below. Results obtained in individual laboratories may differ.

Precision

Precision was determined using Elecsys reagents, human sera and controls (repeatability n = 21, intermediate precision n = 10); intermediate precision on MODULAR ANALYTICS E170 analyzer was determined in a modified protocol (EP5-A) of the CLSI (Clinical and Laboratory Standards Institute): 6 times daily for 10 days (n = 60). The following results were obtained:

Elecsys 2010 and cobas e 411 analyzers						
Sample	Repeatability			Intermediate precision		
	Mean COI ^{d)}	SD COI	CV %	Mean COI	SD COI	CV %
HS ^{e)} , negative	0.109	0.002	2.2	0.103	0.006	5.4
HS, positive	1.37	0.021	1.5	1.33	0.034	2.5
HS, positive	3.78	0.067	1.8	3.70	0.171	4.6
PC ^{f)} Toxo IgM 1	0.120	0.002	1.6	0.118	0.005	4.1
PC Toxo IgM 2	1.35	0.015	1.1	1.29	0.043	3.3

d) COI = cutoff index

e) HS = human serum

f) PC = PreciControl

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MODULAR ANALYTICS E170, cobas e 601 and cobas e 602 analyzers						
Sample	Repeatability			Intermediate precision		
	Mean COI	SD COI	CV %	Mean COI	SD COI	CV %
HS, negative	0.107	0.002	1.8	0.103	0.002	1.9
HS, positive	1.33	0.011	0.9	1.36	0.023	1.7
HS, positive	3.86	0.034	0.9	3.83	0.061	1.6
PC Toxo IgM 1	0.116	0.002	1.6	0.117	0.002	1.7
PC Toxo IgM 2	1.30	0.015	1.2	1.31	0.032	2.4

Method comparison

In study 1 the performance of the Elecsys Toxo IgM assay was determined by testing a total of 826 fresh and frozen samples at two sites in comparison to a commercially available *Toxoplasma* IgM test.

In study 2 the Elecsys Toxo IgM assay was compared to another commercially available *Toxoplasma* IgM assay by testing 400 fresh and frozen samples. In both studies all specimens with initially discordant results were re-tested. Resolution of repeatedly discordant samples was done by avidity testing. 51 specimens with indeterminate results in one of the assays were excluded from the final calculation of relative sensitivity and specificity.

Relative sensitivity and specificity after resolution

Study	N	Relative sensitivity %	Lower confidence limit %	Relative specificity %	Lower confidence limit %
1	785	95.3 (162/170)	91.7	98.9 (595/602)	97.8
2	390	98.8 (83/84)	94.4	99.7 (294/295)	98.4

Study 1: Of 21 samples which were initially discordant negative with the Elecsys Toxo IgM assay, 11 samples revealed a high avidity test result, 2 samples were found negative with Toxo ISAGA IgM. 7 discordant negative samples revealed a low avidity test result, 1 sample was found positive with Toxo ISAGA IgM. 5 samples which were discordant positive with the Elecsys Toxo IgM assay revealed a high avidity test result, 2 samples were from individuals without *Toxoplasma* infection.

Study 2: Of 12 samples which were initially discordant negative with the Elecsys Toxo IgM assay, 11 samples revealed a high avidity test result. 1 sample revealed a low avidity test result. 1 sample which was discordant positive with the Elecsys Toxo IgM assay was from an individual without *Toxoplasma* infection.

Analytical specificity

455 potentially cross reacting samples were tested with the Elecsys Toxo IgM assay and a comparison Toxo IgM assay comprising specimens:

- containing antibodies against HAV, HBV*, HCV, HIV, CMV, EBV*, HSV, VZV, Rubella, *Treponema pallidum*, Malaria**, Amebiasis, Chlamydia and Gonorrhea
- containing autoantibodies (AMA*, ANA) and elevated titers of rheumatoid factors
- after vaccination against HBV and Influenza

An overall agreement of 99.1 % (446/451) was found in these specimens using the Elecsys Toxo IgM assay and the comparison test. 444 samples were found concordantly negative and 2 samples were found positive. 4 samples were found indeterminate either by the Elecsys Toxo IgM assay or the comparison test.

* 1 discordant sample in each of these groups

** 2 discordant samples

Seroconversion panels

In two studies seroconversion samples obtained during pregnancy screening were tested with the Elecsys Toxo IgM assay in comparison to two different commercially available *Toxo* IgM assays.

In 24 seroconversion panels comprising 83 samples at the first site, the Elecsys Toxo IgM assay detected 64 samples from 66 samples which were found positive using a comparison test. 2 discordant negative sera were follow-up samples, taken more than 8 weeks after infection.

In 29 seroconversion panels (including 92 samples) at the second site, the Elecsys Toxo IgM assay detected 67 samples from 74 samples which were found positive by a second comparison test. 2 discordant negative sera from the very early phase of infection were also negative by another comparison test. In two panels (comprising 3 and 2 serial bleeds from the very early phase of infection) IgM was not detected, however seroconversion could be demonstrated by the Elecsys Toxo IgG assay.

In both panels discordant negative results for several samples were also found by two other commercial *Toxoplasma* IgM assays.

References

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For further information, please refer to the appropriate operator's manual for the analyzer concerned, the respective application sheets, the product information and the Method Sheets of all necessary components (if available in your country).

A point (period/stop) is always used in this Method Sheet as the decimal separator to mark the border between the integral and the fractional parts of a decimal numeral. Separators for thousands are not used.

Symbols

Roche Diagnostics uses the following symbols and signs in addition to those listed in the ISO 15223-1 standard.

CONTENT	Contents of kit
SYSTEM	Analyzers/Instruments on which reagents can be used
REAGENT	Reagent
CALIBRATOR	Calibrator
→	Volume after reconstitution or mixing
GTIN	Global Trade Item Number

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